Introduction

Flt3 receptor gene is an extracellular receptor and primitive hematopoietic progenitor, which is belonged to receptor tyrosine kinase III (RTKIII). It is comprised of five extracellular immune-globin like (Ig-Like) regions and one intracellular kinase region and these two are connected through a permeable membrane. Flt3 receptor gene is a 993 amino acid protein [1, 2]. This gene is positioned on the short arm of chromosome 13 (13q12,2) and expressed in immature hematopoietic cell, placenta, gonad, and the brain [3]. Flt3 ligand (FL) is also expressed in most of cell lines [4-5]. Normally and despite stimulation by flt3 receptors, even ligands cannot induce growth independently from this cells, while it can induces proliferation independent from IL-3 factor of cell in mutated form like ITD or point mutation (D835)[6]. Despite considerable variation along amino acids replication at the range of 3-50 amino acids, there is no difference in biological activity of flt3/ITD alleles in cell culture or animal models [7-8]. Cooperation between flt3 mutation and genetic rearrangement may be needed for development of AML phenotype.

Excessive expression of flt3 reception gene happens in 70-100% of AML and notable percentage of ALL [9-16]. Mutation in flt3 receptor gene is featured with largest frequency of genetic disorders in AML and the most renowned mutation is ITD with mutation point at ASP835 [10-11].

ITD mutation occurs in exon 11 & 12 and intron 11 and the first case of ITD at juxtamembrane (JM) was reported by Nakao et al. (1996). This mutation represents about 20-25% of AML cases. D835 point mutation also occurs in exon 17 of flt3 receptor gene and at aspartic acid[10-11].

Methods

Totally, 91 children with acute myeloid leukemia (AML) and acute lymphoid leukemia (ALL) were examined as to flt3 mutation, internal tandem duplication (ITD) mutation, and point mutation in exon 17 (e17). ITD mutation in flt3 receptor was carried out in exon 12&12 and intron 11. As to point mutation in e17, PCR products of the subjects after PCR on genomic DNA of them were examined using restriction enzyme (ECORV) and restriction fragment length polymorphism (RFLP). As to ITD positive, sequencing method was used. Findings: ITD mutation was observed in seven (7.7%) of acute leukemia patients and two (2.2%) patients were diagnosed with point mutation D835. Distribution in different subgroups of FAB was not identical. Conclusion: FLT3 mutation was highly prevalent in children with acute leukemia. Therefore, molecular diagnosis of these mutation, regardless of FAB categorization and before initiation of intervention, can be used to make better decision about therapeutic protocol.

Keywords: Acute leukemia- FLT4 gene- ITD mutation- point mutation D835
acid 835-amino acid. Substitution ASP835Tyr is the most common case [12]. As recommended by the data, about 30% of AML patients demonstrate acquired mutation in flt3 receptor gene, which consists of 24% ITD and 6-7% point mutation at aspartic acid 835. Total frequency of the mutation in the adults with AML is 24% and this figure in children ranges from 10% to 15%. In other words, frequency of the mutation increases with age and frequency of ITD in ALL is lower and about 1-5%. Therefore, and given the statistics of mutation of other genes, flt3 receptor is the most common mutated gene in AML patients [13].

Given the importance of mutation prognosis value of flt3 receptor gene in AML patients, a great deal of attention has been drawn to development of drugs to suppress flt3 activity.

Taking into account paucity of studies on this type of mutation in Iran, it is necessary and essential to develop methods for screening activation mutation, implement molecular diagnosis methods, and survey expression frequency of the mutation in acute leukemia patients along with the mutations observed in tyrosine kinases or other downstream factors effective on the mutation in acute leukemia cases.

Materials and Methods

Mutation of flt3 receptor gene in 91 children with AML with different FAB categorization groups (AML-M3 in particular) and ALL with immunophenotype categorization were examined through flow cytometer as a basic study. Blood samples were supplied from blood ward of Aliasghar Hospital, Bahrami Hospital, Children Clinic, and flow cytometer center of Iran Blood Transfusion Organization. Extraction was done through proteinase K and saturated ammonium acetate. Blood vails were kept in a freezer until DNA extraction. To diagnose ITD mutation genomic DNA was extracted by Forward Primer5'-GCAATTTAGGTATGAAAGCCAGC-, reverse primer5'-CTTTTTTGACGGCAACCTGCA-, and initial denaturation 1x based on PCR program (95ºC, 5min), annealing (66ºC, 1min), denaturation (95ºC, 1min), 35x elongation (72ºC, 90s), and final elongation (72ºC, 7min). Mutation cases were detected using direct observation and comparing bands and markers.

As to point mutation D835, reverse primer5'-GCAGCCTCACATTGCGCAAGGCCACGC-, forward primer 5'-CCGGCCA GGA ACG TGC TTG-3', and initial denaturation 1x following 1x PCR program (95ºC, 5min), annealing (60ºC, 1min), denaturation (95ºC, 1min), 35x elongation (72ºC, 96sec), and final elongation (72ºC, 7min) were used on the cultured genome extracted from the patients and mastermix including 5µ buffer, 1 µL dNTP, 1.5 µL MgCl2, 10 µL, 1 µLR primer, and 1 µLF primer. To detect point mutation D835 based on RFLP technique, ECORV enzyme was placed in incubator (16hrs, 37ºC). Marking was done in ethidium bromide, the samples were rinsed by distilled water and inspected before imaging. The bands indicated whether there was a mutation or not. In ITD positive cases, sequencing method was used for confirmation.

For sequencing, the pieces with PCR mutation were purified using the primers and five samples were sequenced using ABI 3130 sequencing set. The diagrams were read using Chromas and the insertion mutations were spotted through comparing them with non-mutated sample sequences.

Results

Among 91 patients under study, seven patients (7.7%) showed ITD mutation, so that out of one M0, none was observed in none them; out of three M1, one was observed in one subject; out three M2, one was observed in one subject; out of two M3, two were observed in two subjects; out of three M4, none was observed; out of two M2, none was observed among them; out of three M6, none was observed among them; out of 39 Early pre B cells, three were observed in three of them; out of 14 Pre B cell, none was observed among them; out of five B cell, none was observed in them, and out of 15 T cell cases, none was observed in them. (Figure 1)

As indicated by the marker in Figure 1, pieces 328bp are observable, which are pertinent to exon 11 and 12 and intron 11 of flt3 gene. The above samples belong to different sub-categories of FAB category and among them, column two represents AMP patients-M3; columns six and eight represent Early pre B cell patient with mutation; column nine represents patient with AML-M2; and column 13 represents AML-M1 patient. In

Figure 1. ITC Mutation in Flt3 Gene in Different Subgroups of Acute Leukemia Patients; N, Negative Control; M, Marker

Figure 2. D835 Mutation in Flt3 Gene in Different Subgroups of AML And ALL Patients; N, Negative Control; M, Marker
Table 2. Information of Leukemic Patients with ITD and D835 Mutations

<table>
<thead>
<tr>
<th>Age (year)</th>
<th>Gender</th>
<th>Disease</th>
<th>Karyotype</th>
<th>ITD</th>
<th>D835</th>
</tr>
</thead>
<tbody>
<tr>
<td>9</td>
<td>F</td>
<td>AML/M2</td>
<td>Unknown</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>17</td>
<td>F</td>
<td>Early pre Bcell</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>F</td>
<td>Early pre Bcell</td>
<td>Normal</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>15</td>
<td>M</td>
<td>AML/M1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>M</td>
<td>AML/M3</td>
<td>Normal</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>13</td>
<td>M</td>
<td>Early pre Bcell</td>
<td>48, XY, +8 (t(15;17)(q22; q21), +21 [15]/46, XY[5])</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>4</td>
<td>M</td>
<td>AML/M3</td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>F</td>
<td>Early Pre Bcell</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Discussion

ITD mutation was observed in seven patients (7.7% of the study group) with acute leukemia and 2 patients (2.2%) showed point mutation D835 so that distribution of them in different FAB subgroups was not identical. Positive cases of ITD mutation was observed in M1, M2, M3, and Early pre B cell.

The mutation under study is highly important for leukemia pathogens. Mutation in flt3 receptor belonged to RTKII and induced uncontrolled proliferation of leukemia cells. Studies with larger sample groups have reported ITD mutation with bad prognosis and development of leukemia that resists standard treatments [15].

Bad prognosis has been reported by three studies on children with AML and flt3 mutation. The study with largest sample group (n = 91) by CCG group reported 15 cases of flt3/ITD mutation in 91 AML patients (16.5%). Regress of the disease in flt3/ITD+ and flt3/ITD+ patients was 40% and 73% respectively (p = 0.005). Event free survival (EFS) of eight years was reported in 7% and 44% of flt3/ITD+ and flt3/ITD+ patients respectively (p = 0.002). Highest frequency was observed in the subgroup M3. Flt3/ITD mutation in adults usually refers to a bad prognosis [14-17].

The results showed flt3/ITD mutations in 23.5% of AML and 4% of ALL children. Regardless of number of insertion or duplicate nucleotide, presence of amino acid of whatever number causes spatial deformation of the protein so that preventive effect of JM region on auto-phosphorylation fades away. The duplications can occur in exon 11 and 12 and intron 11 of flt3 gene to be inserted in the common border of exon and intron 11 [15-18]. Point mutation in exon 17 of flt3 receptor gene was also examined, where aspartic acid 835 – amino acid (D835) is substituted by other amino acids such as tyrosine, valine, etc.. The substitution leads to development of activation loop. Among 91 patients with acute leukemia, two patients demonstrated the mutation, which was AML in 6% of the cases and ALL in 1.4% of the cases. Due to low rate of the mutation and since the sample group was small, it is not possible to define a relationship between the mutation and FAB subcategories. A study reported ASP835 substitution in 30 subjects among 429 AML children (7%), one subject among 29 MDS patients (3%), and one subject among 36 ALL patients (3%). A similar study reported the mutation in seven subjects among 35
AML patients (7%). Occurrence of point mutation D835 was significantly less than ITD mutation in flt3 gene; while the both mutations were highly frequent in AML. Both ITD and D835 mutations in flt3 gene were reported in a patient; however sequencing studies have shown that these two mutations were not on the same allele.

Here, ITD mutation was observed in 7.7% of the patients without identical distribution among FAB subcategories for leukemia categorization. As suggest by studies, flt3/ITD mutation has been observed in 10-20% of the children with AML and 1-5% with ALL. In general, the mutation was significantly associated with weak emergence and bad prognosis in ALL and AML patients.

As to point mutation D in flt3 gene, 4% of the ALL and AML patients demonstrated such a mutation. This figure in other studies is reported 5% with different distribution in different FAB subcategories. Effective follow up of ITD and D835 mutation in flt3 receptor gene and carrying out molecule diagnosis as a routine procedure will lead to better decisions as to treatment of AML and ALL and avoid development of resistance and frequent recurrences.

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References